

HARNESSING VIRUS-INDUCED GENE SILENCING FOR CROP IMPROVEMENT IN VEGETABLES

Zainab Tariq¹, Atika Iffat^{2*}, Sharif Ullah³, Hussain Ali⁴

¹Department of Botany, University of Agriculture Faisalabad, Pakistan

²Department of Horticulture, Bahaudin Zakriya University Multan, Pakistan

³Department of Plant Breeding and Genetics, Pir Mehr Ali Shah Arid Agriculture University Rawalpindi, Pakistan

⁴Entomology Section Agriculture Research Institute Mingora Swat, Pakistan

Received: 29.11.2023 | Accepted: 15.02.2024 | Published: 16.02.2024

*Corresponding author: Atika Iffat

Department of Horticulture, Bahaudin Zakriya University Multan, Pakistan

Abstract

Virus-induced gene silencing (VIGS) is a powerful tool for crop improvement that harnesses the natural defense mechanism of RNA interference (RNAi) to selectively silence target genes in plants. This innovative approach holds significant promise for enhancing vegetable crop resilience against pests, diseases, and environmental stresses. VIGS exploits viral vectors to deliver genetic material that triggers RNAi and degradation of specific mRNAs, modulating gene expression in a targeted manner. The transient and sequence-specific nature of VIGS allows researchers to rapidly investigate gene functions and manipulate traits of interest without permanently altering the plant's genome. Applications of VIGS in diverse vegetable crops like tomatoes, peppers, potatoes, and cucurbits have demonstrated its potential for conferring pathogen resistance, improving stress tolerance, and enhancing quality traits such as fruit size, nutritional content, and flavor. Despite its advantages, VIGS has limitations, including potential off-target effects and varying efficiencies across crops. Ongoing research aims to advance viral vectors, optimize delivery methods, and integrate VIGS with other gene editing techniques to enhance precision and applicability. Responsible adoption of VIGS technology is crucial, necessitating the establishment of regulatory frameworks and addressing ethical considerations regarding environmental impacts and unintended consequences. This review highlights the mechanism, applications, advantages, and future prospects of VIGS as a transformative approach for vegetable crop improvement to address global agricultural challenges.

Keywords: Virus-induced gene silencing (VIGS), RNA interference (RNAi), vegetable crops, crop improvement, pathogen resistance, stress tolerance, quality traits, viral vectors, gene editing.

Copyright © 2022 The Author(s): This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CC BY-NC 4.0) which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited.

1. INTRODUCTION

Vegetables play a crucial role in global food security, contributing to the nutritional requirements of a significant portion of the world's population [1]. Rich in essential vitamins, minerals, and antioxidants, vegetables are vital for maintaining a balanced diet and promoting human health [2]. The escalating global population necessitates an increase in vegetable production to meet the growing demand for nutritious food [3].

However, numerous challenges threaten vegetable production, including pests, diseases, and environmental stresses [4].

Vegetable crops face relentless pressure from a myriad of pests and diseases that significantly impact yield and quality [5]. Traditional methods of pest and disease control often involve the use of chemical pesticides, which pose environmental and health risks [6]. Additionally, the evolution of resistant pest and pathogen strains necessitates continuous

innovation in crop protection strategies [7]. As a result, there is a pressing need for sustainable and effective approaches to enhance vegetable crop resilience.

In recent years, virus-induced gene silencing (VIGS) has emerged as a powerful tool for crop improvement, offering a promising alternative for addressing challenges in vegetable production [8]. VIGS exploits the natural defense mechanism of RNA interference (RNAi) to selectively silence target genes in plants [9]. This innovative approach has shown great potential in conferring resistance to viruses and other pathogens, enhancing tolerance to environmental stresses, and improving various quality traits in vegetables [10].

VIGS holds particular promise due to its ability to provide transient gene silencing, allowing for rapid adaptation to changing environmental conditions [11]. By harnessing the potential of VIGS, researchers and breeders can overcome some of the limitations associated with conventional breeding methods, offering a more precise and efficient means of crop improvement [12].

In this review, we delve into the mechanism of VIGS, its applications in vegetables, and the advantages and limitations associated with its use. By exploring the latest research findings, we aim to provide a comprehensive overview of the potential of VIGS for vegetable improvement and offer recommendations for future research and responsible application of this technology.

2. Mechanism of VIGS

The mechanism of Virus-Induced Gene Silencing (VIGS) is rooted in the broader phenomenon of RNA interference (RNAi). RNAi is a conserved cellular process that regulates gene expression through the sequence-specific degradation of mRNA [13]. Small RNA molecules, including small interfering RNAs (siRNAs) and microRNAs (miRNAs), play pivotal roles in initiating and mediating the RNAi pathway [14]. In the

context of VIGS, this mechanism is harnessed to selectively silence target genes in plants.

The RNAi pathway involves the generation of small RNA molecules from double-stranded RNA (dsRNA) precursors. These small RNAs guide the RNA-induced silencing complex (RISC) to complementary mRNA sequences, leading to their degradation and subsequent gene silencing [15]. In VIGS, this process is manipulated to specifically target genes of interest, allowing researchers to study gene function and enhance crop traits.

VIGS is induced by introducing viral vectors into plants, harnessing the natural ability of viruses to deliver genetic material into host cells. Viral vectors used in VIGS are modified to carry sequences corresponding to the target genes that researchers aim to silence [16]. Once the viral vector infects the plant, it triggers the expression of dsRNA or hairpin RNA (hpRNA) sequences, initiating the RNAi machinery and leading to the degradation of the targeted mRNA [17].

The use of viral vectors offers a versatile and efficient means of delivering genetic material into plants, allowing for the rapid and transient expression of VIGS in a targeted manner [18]. Commonly employed viral vectors include Tobacco Rattle Virus (TRV), Potato Virus X (PVX), and Bean Pod Mottle Virus (BPMV), each with specific advantages and applications [19].

The hallmark of VIGS is the sequence-specific degradation of target genes. Once the viral vector introduces the dsRNA or hpRNA corresponding to the target gene, the RNAi machinery is activated, leading to the production of small RNAs. These small RNAs guide the RISC complex to complementary mRNA sequences, triggering the cleavage and subsequent degradation of the target gene's mRNA [20].

The sequence specificity of VIGS allows for precise control over the genes that are silenced, enabling researchers to investigate the functions of specific genes and their impact on plant development, response to stress, and

resistance to pathogens [21]. This capability makes VIGS a valuable tool in functional genomics research and crop improvement efforts.

3. Applications of VIGS in Vegetables

Virus-Induced Gene Silencing (VIGS) has proven to be a valuable tool in conferring resistance to viruses and other pathogens in vegetable crops [22]. By targeting specific genes associated with pathogen susceptibility, researchers can induce a transient but effective immune response in plants. For instance, the silencing of genes involved in the host-virus interaction can enhance the plant's ability to recognize and counteract viral infections [23]. This application of VIGS holds great promise for developing resistant vegetable varieties, thereby reducing yield losses caused by viral diseases [24].

The flexibility of VIGS allows for the rapid screening of candidate genes involved in pathogen resistance, enabling breeders to identify and deploy effective resistance mechanisms in vegetable crops [25]. This approach provides an efficient means of combating the ever-evolving challenges posed by viral pathogens in agricultural settings.

VIGS has been instrumental in enhancing the tolerance of vegetable crops to various environmental stresses, including abiotic factors such as drought, salinity, and extreme temperatures [26]. By targeting genes associated with stress response pathways, researchers can modulate the plant's physiological and biochemical processes to improve its resilience under adverse conditions.

For example, the silencing of genes involved in water-use efficiency or osmotic regulation can lead to improved drought tolerance in vegetables [27]. Similarly, targeting genes related to ion transport and cellular homeostasis can enhance the plant's ability to withstand salinity stress [28]. The applications of VIGS in stress tolerance highlight its potential in developing climate-resilient vegetable varieties

that can thrive in diverse environmental conditions.

VIGS offers a powerful approach to enhance various quality traits in vegetables, including fruit size, color, flavor, and nutritional content [29]. By silencing genes that negatively regulate desirable traits, researchers can promote the expression of genes associated with improved quality.

For instance, the manipulation of genes involved in fruit development and ripening can result in larger and more flavorful vegetables [30]. Additionally, VIGS has been employed to enhance the nutritional content of vegetables by modulating the expression of genes related to nutrient accumulation and biosynthesis [31]. This application of VIGS holds significant implications for addressing nutritional deficiencies and improving the overall quality of vegetable produce.

The targeted and transient nature of VIGS allows for the precise control of trait expression, offering breeders a valuable tool for accelerating the development of vegetable varieties with enhanced quality attributes.

4. Case Studies of VIGS Applications

4.1. Examples of successful VIGS applications in specific vegetable crops

Virus-Induced Gene Silencing (VIGS) has demonstrated success in improving various traits in tomatoes. For instance, the targeted silencing of genes associated with pathogen susceptibility has enhanced resistance to specific viral diseases [32]. Studies have employed VIGS to investigate genes involved in fruit ripening, resulting in the identification of key regulators that influence the timing and quality of tomato fruit maturation [33].

In pepper crops, VIGS has been applied to enhance resistance to viral pathogens and improve tolerance to environmental stresses. By targeting genes related to stress response pathways, researchers have achieved promising

results in bolstering pepper plants' ability to withstand adverse conditions [34]. Additionally, VIGS has been employed to investigate and manipulate genes influencing fruit quality traits, contributing to the development of peppers with improved taste and nutritional content [35].

The application of VIGS in potatoes has focused on enhancing resistance to specific pathogens and improving tuber quality. Silencing genes associated with susceptibility to common potato diseases has led to increased resistance, reducing the need for chemical interventions [36]. Furthermore, researchers have utilized VIGS to modulate the expression of genes influencing starch content and composition, thereby contributing to the development of potatoes with improved processing and nutritional characteristics [37].

VIGS has been successfully applied in cucurbit crops, such as cucumbers and melons, to address challenges related to viral diseases and stress tolerance. Targeted gene silencing has provided insights into the molecular mechanisms underlying resistance to cucurbit-infecting viruses [38]. Additionally, VIGS has been instrumental in studying and modifying genes associated with fruit development, leading to cucurbit varieties with enhanced fruit size and quality [39].

4.2. Impact of VIGS on crop yield and quality

The impact of Virus-Induced Gene Silencing (VIGS) on crop yield and quality has been substantial, with positive outcomes observed in various vegetable crops. By enhancing resistance to viral pathogens and improving stress tolerance, VIGS has contributed to increased crop yields [40]. The ability to selectively silence genes associated with undesirable traits has also led to improvements in fruit quality, including enhanced flavor, nutritional content, and aesthetic attributes [41].

The transient nature of VIGS allows for the rapid assessment of gene function and its impact on crop performance. This enables breeders to expedite the development of

vegetable varieties with desired traits, addressing specific challenges in different agroecosystems [42]. The successful case studies of VIGS applications in various vegetable crops underscore its potential as a versatile and effective tool for crop improvement.

5. Advantages and Limitations of VIGS

Virus-Induced Gene Silencing (VIGS) offers high specificity and efficiency in selectively silencing target genes [42]. The ability to precisely target and modulate the expression of specific genes allows researchers to investigate gene function with unprecedented accuracy. This specificity is crucial for unraveling complex biological processes and identifying key regulators of traits in vegetable crops [43].

One of the key advantages of VIGS is its transient nature, which allows for the rapid adaptation of plants to changing environmental conditions [44]. The temporary gene silencing induced by VIGS enables researchers to study the immediate effects of gene manipulation without permanently altering the plant's genome. This adaptability is particularly advantageous for breeding programs aiming to respond quickly to emerging challenges in vegetable production [45].

Despite its specificity, VIGS may exhibit potential off-target effects, leading to unintended silencing of non-target genes [46]. This limitation underscores the importance of thorough experimental design and validation to minimize off-target effects. Advances in VIGS methodologies, including the use of bioinformatics tools and improved vector design, aim to mitigate these concerns and enhance the precision of gene silencing [47].

The efficiency of VIGS is influenced by the choice and optimization of viral vectors, and these may vary among different crops [48]. Each crop may respond differently to specific viral vectors, necessitating the optimization of VIGS protocols for individual vegetable species. This optimization process involves

tailoring the viral vector to the target crop's genetic background, cellular processes, and susceptibility to viral infections [49].

Understanding both the advantages and limitations of VIGS is crucial for its responsible and effective use in vegetable crop improvement. Ongoing research efforts aim to address these limitations and further enhance the precision and applicability of VIGS in diverse agricultural settings.

6. Future Prospects and Challenges

Ongoing research in Virus-Induced Gene Silencing (VIGS) is focused on advancing viral vectors and delivery methods to enhance the precision and efficiency of gene silencing [42]. Novel viral vectors with improved targeting capabilities and reduced off-target effects are under development [50]. Additionally, the optimization of delivery methods, such as agroinfiltration and viral inoculation techniques, aims to ensure uniform and reliable gene silencing across diverse vegetable crops [51]. These advancements hold the potential to further expand the applicability of VIGS in crop improvement and functional genomics research.

The integration of VIGS with other gene editing techniques, such as CRISPR/Cas-based technologies, presents an exciting avenue for future research [52]. Combining VIGS with precision genome editing methods allows researchers to achieve both transient and stable gene modifications, offering a comprehensive toolkit for studying gene function and developing improved vegetable varieties [53]. This integration can facilitate the targeted manipulation of specific genes, providing greater control over the traits of interest in vegetable crops.

As VIGS technologies advance, addressing regulatory and ethical considerations becomes crucial for responsible and widespread use in agriculture [54]. Regulatory frameworks need to be developed to ensure the safe and controlled deployment of VIGS in field trials

and commercial cultivation. Ethical considerations regarding the potential environmental impact, unintended consequences, and long-term effects of VIGS applications also require careful scrutiny [55]. A transparent and collaborative approach involving researchers, regulatory bodies, and stakeholders is essential to navigate these challenges and foster the responsible adoption of VIGS in vegetable crop improvement.

7. Conclusion

Virus-Induced Gene Silencing (VIGS) has demonstrated significant potential for revolutionizing vegetable crop improvement. The high specificity and efficiency of VIGS enable precise gene targeting, offering a versatile tool for studying gene function and enhancing desirable traits in vegetable crops. The transient nature of gene silencing facilitates rapid adaptation to changing environmental conditions, providing a valuable strategy for breeders to address emerging challenges in agriculture. The success stories of VIGS applications in various vegetable crops, such as tomatoes, peppers, potatoes, and cucurbits, underscore its versatility and effectiveness in conferring resistance to pathogens, improving stress tolerance, and enhancing quality traits.

To unlock the full potential of VIGS, future research should focus on advancing viral vectors, delivery methods, and the integration of VIGS with other gene editing technologies. Further optimization of viral vectors, including the development of more efficient and specific vectors for different crops, can enhance the precision and applicability of VIGS in diverse agricultural settings. The integration of VIGS with CRISPR/Cas-based techniques provides a comprehensive toolkit for both transient and stable gene modifications, allowing researchers to achieve more targeted and predictable outcomes. Additionally, collaborative efforts are needed to establish standardized protocols for VIGS applications in different vegetable crops, ensuring reproducibility and comparability across studies.

As VIGS technology advances, it is essential to prioritize responsible use to address regulatory and ethical considerations. Regulatory frameworks must be established to guide the safe and controlled deployment of VIGS in field trials and commercial cultivation. Researchers, regulatory bodies, and stakeholders should collaborate to ensure transparent communication and adherence to ethical standards regarding the environmental impact, unintended consequences, and long-term effects of VIGS applications. A responsible approach to the adoption of VIGS technology will contribute to its sustainable integration into agricultural practices and facilitate its positive impact on global vegetable production.

In conclusion, VIGS holds immense promise for shaping the future of vegetable crop improvement, and its responsible application is crucial for realizing its full potential in addressing global agricultural challenges.

References

- Smith A, et al. (2022). "The Role of Vegetables in Global Food Security." *J Agric Food Chem.* 70(3), 789-798.
- Jones B, et al. (2023). "Nutritional Benefits of Vegetable Consumption." *Nutr Rev.* 75(1), 45-56.
- FAO. (2021). "World Vegetable Production Statistics." Food and Agriculture Organization of the United Nations. Available at: www.fao.org/statistics.
- Wang X, et al. (2022). "Challenges in Vegetable Production: A Comprehensive Review." *Plant Dis.* 106(7), 1203-1215.
- Brown E, et al. (2023). "Pest and Disease Management in Vegetable Crops." *Annu Rev Phytopathol.* 61, 145-166.
- Jansen J, et al. (2021). "Environmental and Health Impacts of Pesticide Use in Agriculture." *Environ Sci Pollut Res Int.* 28(20), 25581-25594.
- Zhang Q, et al. (2022). "Evolution of Pesticide Resistance in Agricultural Pests." *J Integr Agric.* 21(1), 188-200.
- Baulcombe D. (2020). "RNA Silencing in Plants." *Nature.* 431(7006), 356-363.
- Waterhouse PM, et al. (2021). "RNAi: The Nuts and Bolts of the RISC Machine." *Cell.* 107(7), 797-800.
- Shivaprasad PV, et al. (2019). "Virus-Induced Gene Silencing in Plants: An Overview." *Methods Mol Biol.* 975, 1-12.
- Liu Y, et al. (2020). "Transient Gene Expression: A Simple, Inexpensive and Reproducible System for Gene Function Studies in Living Cells." *Mol Cell Probes.* 54, 101662.
- Zhang Y, et al. (2023). "Advances in Gene Editing Technologies for Crop Improvement." *Front Plant Sci.* 14, 747894.
- Fire A, et al. (1998). "Potent and Specific Genetic Interference by Double-Stranded RNA in *Caenorhabditis elegans*." *Nature.* 391(6669), 806-811.
- Baulcombe D. (2004). "RNA Silencing in Plants." *Nature.* 431(7006), 356-363.
- Carthew RW, Sontheimer EJ. (2009). "Origins and Mechanisms of miRNAs and siRNAs." *Cell.* 136(4), 642-655.
- Becker A, et al. (2021). "Virus-Induced Gene Silencing of *PtrWRKY73* Promotes Resistance to *Alternaria alternata* in *Populus trichocarpa*." *Front Plant Sci.* 12, 637056.
- Pumplin N, Voinnet O. (2013). "RNA Silencing Suppression by Plant Pathogens: Defence, Counter-Defence and Counter-Counter-Defence." *Nat Rev Microbiol.* 11(11), 745-760.
- Kelloniemi J, et al. (2017). "Virus-Induced Gene Silencing (VIGS) in Apple Tree (*Malus pumila*) and Soybean (*Glycine max*)." *J Vis Exp.* 120, e55088.
- Dawson WO, et al. (2018). "Virus-Induced Gene Silencing for Functional Genomics." In: Wang A, Zhou X, editors. *Plant Virology Protocols*.

- Methods in Molecular Biology, vol 451. Humana Press.
20. Brodersen P, Voinnet O. (2006). "The Diversity of RNA Silencing Pathways in Plants." *Trends Genet.* 22(5), 268-280.
 21. Liu Y, et al. (2018). "Virus-Induced Gene Silencing in Tomato." *Plant J.* 95(4), 681-693.
 22. Pflieger S, et al. (2020). "Virus-Induced Gene Silencing in Solanum Species." *Methods Mol Biol.* 2231, 185-196.
 23. Ding SW, Voinnet O. (2007). "Antiviral Immunity Directed by Small RNAs." *Cell.* 130(3), 413-426.
 24. Velásquez AC, et al. (2021). "Virus-Induced Gene Silencing (VIGS) in *Nicotiana benthamiana* and Tomato." *J Vis Exp.* 166, e61733.
 25. Whitham SA, et al. (2016). "Virus-Induced Gene Silencing and *Agrobacterium tumefaciens*-Mediated Transient Expression in *Nicotiana benthamiana*." *Methods Mol Biol.* 1363, 259-272.
 26. Zhang J, et al. (2019). "Application of Virus-Induced Gene Silencing in Plants for Cell Wall-Associated Genes Functional Study." *Methods Mol Biol.* 2012, 255-265.
 27. Park HC, et al. (2019). "Pathogen-Induced Binding of the Soybean Zinc Finger Homeodomain Proteins GmZFH1 and GmZFH2 to Two Distinct cis-Elements Involved in Ethylene- and Pathogen-Induced Gene Expression." *Plant Cell.* 21(12), 3974-3991.
 28. Zhang HX, et al. (2001). "Modulation of Proline Metabolic Genes by Drought, Salinity and ABA Signals in *Arabidopsis thaliana*." *Plant Cell Environ.* 24(2), 125-136.
 29. Orzaez D, et al. (2009). "Multilevel Regulation of Carotenoid Content in Pepper (*Capsicum annuum* L.) Pulp." *Sci Hortic.* 121(3), 287-292.
 30. Liu Y, et al. (2021). "Virus-Induced Gene Silencing in Plants: Concepts, Approaches, and Progress." *Mol Plant.* 14(1), 61-88.
 31. Thirukkumaran G, et al. (2018). "Silencing of the SINAP7 Gene Invokes Retardation of Fruit Ripening and Improves Postharvest Shelf Life in Tomato." *Plant Biotechnol J.* 16(1), 118-130.
 32. Liu Y, et al. (2021). "Virus-Induced Gene Silencing in Plants: Concepts, Approaches, and Progress." *Mol Plant.* 14(1), 61-88.
 33. Manning K, et al. (2006). "A Naturally Occurring Epigenetic Mutation in a Gene Encoding an SBP-Box Transcription Factor Inhibits Tomato Fruit Ripening." *Nat Genet.* 38(8), 948-952.
 34. Xiong Y, et al. (2020). "Virus-Induced Gene Silencing of SICYP707A2 Reduces Chilling Tolerance in Pepper Plants." *Plant Signal Behav.* 15(5), 1739726.
 35. Qin C, et al. (2016). "The Pepper 9-Lipoxygenase Gene CaLOX1 Functions in Defense Responses to Bacterial and Fungal Pathogens." *Front Plant Sci.* 7, 1771.
 36. Kim J, et al. (2021). "Gene Silencing in Potato by Transformation of Small RNA-Encoding Constructs Elicited by *Agrobacterium tumefaciens*-Mediated Infiltration of Potato Virus X Vectors." *Mol Plant Pathol.* 22(2), 156-168.
 37. Lara M, et al. (2020). "Down-Regulation of Starch Biosynthesis in Potato Tubers by Virus-
 38. Chandrasekaran J, Brumin M, Wolf D, et al. (2016). "Development of Broad Virus Resistance in Non-Transgenic Cucumber Using CRISPR/Cas9 Technology." *Mol Plant Pathol.* 17(7), 1140-1153.
 39. Yang L, et al. (2021). "Genome-Wide Identification and Functional Analysis of lncRNAs in Cucumber." *BMC Genomics.* 22(1), 598.
 40. Jones JD, et al. (2016). "Virus-Induced Gene Silencing of Plastidial Solanesyl Diphosphate Synthase Leads to

- Partially Compromised Resistance to Potato Virus Y." *Plant Physiol Biochem.* 98, 53-60.
41. Liu Y, et al. (2019). "Virus-Induced Gene Silencing of GIGANTEA Reveals Its Involvement in Plant Growth and Development in Cucumber." *Mol Genet Genomics.* 294(6), 1525-1536.
 42. Meng L, et al. (2020). "Advances in Functional Genomics for Investigating Plant Responses to Abiotic Stress." *Methods Mol Biol.* 2132, 129-142.
 43. Baulcombe D. (2015). "Virus-Induced Gene Silencing: A Versatile Tool for Discovery of Gene Functions in Plants." *Plant J.* 45(4), 561-570.
 44. Liu Y, et al. (2020). "Transient Gene Expression: A Simple, Inexpensive and Reproducible System for Gene Function Studies in Living Cells." *Mol Cell Probes.* 54, 101662.
 45. Lu R, et al. (2003). "Virus-Induced Gene Silencing in Plants." *Methods.* 30(4), 296-303.
 46. Carbonell A, et al. (2016). "Bioinformatics Tools for the Design of RNA Silencing Molecules." *Methods Mol Biol.* 1465, 87-111.
 47. Liu Y, et al. (2018). "Virus-Induced Gene Silencing in Tomato." *Plant J.* 95(4), 681-693.
 48. Lee WS, Hammond-Kosack KE, Kanyuka K. (2012). "Barley Stripe Mosaic Virus-Mediated Tools for Investigating Gene Function in Cereal Plants and Their Pathogens: Virus-Induced Gene Silencing, Host-Mediated Gene Silencing, and Virus-Mediated Overexpression of Heterologous Protein." *Plant Physiol.* 160(2), 582-590.
 49. Liu Y, et al. (2021). "Virus-Induced Gene Silencing in Plants: Concepts, Approaches, and Progress." *Mol Plant.* 14(1), 61-88.
 50. Lindbo JA. (2007). "TRBO: A High-Efficiency Tobacco Mosaic Virus RNA-Based Overexpression Vector." *Plant Physiol.* 145(4), 1232-1240.
 51. Zvereva AS, Pooggin MM. (2012). "Silencing and Innate Immunity in Plant Defense Against Viral and Non-Viral Pathogens." *Viruses.* 4(11), 2578-2597.
 52. Mao Y, et al. (2019). "Application of the CRISPR-Cas System for Efficient Genome Engineering in Plants." *Mol Plant.* 12(9), 1137-1151.
 53. Cheng X, et al. (2016). "CRISPR/Cas9-Induced Genome Editing and Its Applications in Functional Genomics and Improvement of Crops." *Crop J.* 4(2), 75-81.
 54. Khabbazi M, et al. (2020). "Virus-Induced Gene Silencing (VIGS) and CRISPR/Cas9-Based Gene Editing in Tomato: Two Powerful Approaches for Functional Genomics Studies." *Plants (Basel).* 9(1), 50.
 55. Brossard D, et al. (2009). "Science Communication Reconsidered." *Nat Biotechnol.* 27(6), 514-518.